

# Immobilization and catalytic properties of pumpkin seed lipase on chitosan

---



Pimwouth  
Natisoontorn



Jiratt Stephen  
Lumpaopong



Sudthipat  
Jithaeng

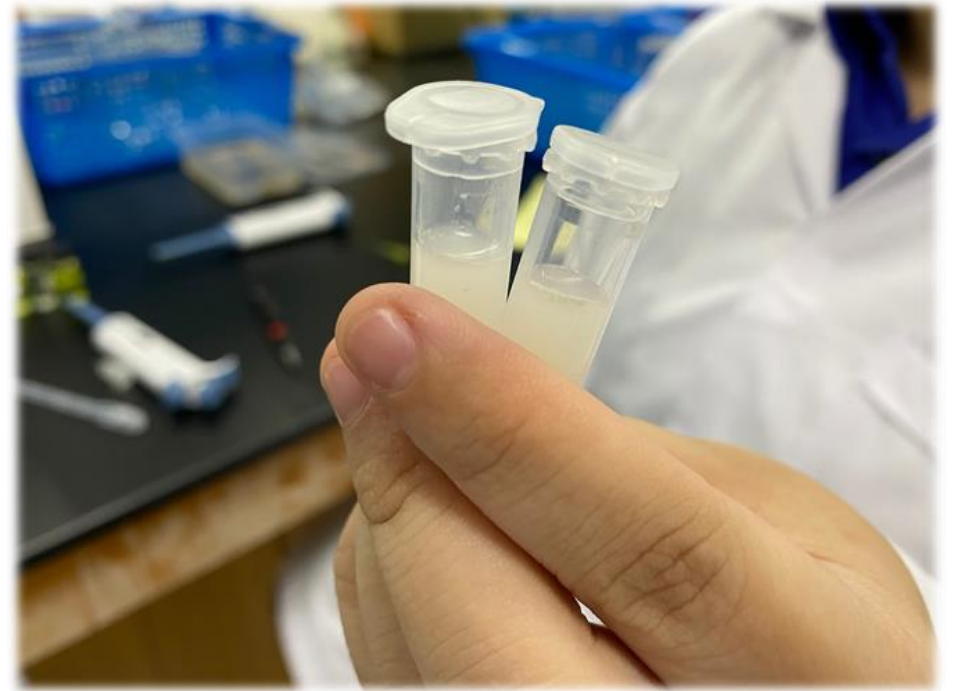


Advisor : Dr. Duangkhae Srikun

Mahidol Wittayanusorn School

# Outline

- Introduction
- Methodology
- Results
- Conclusion

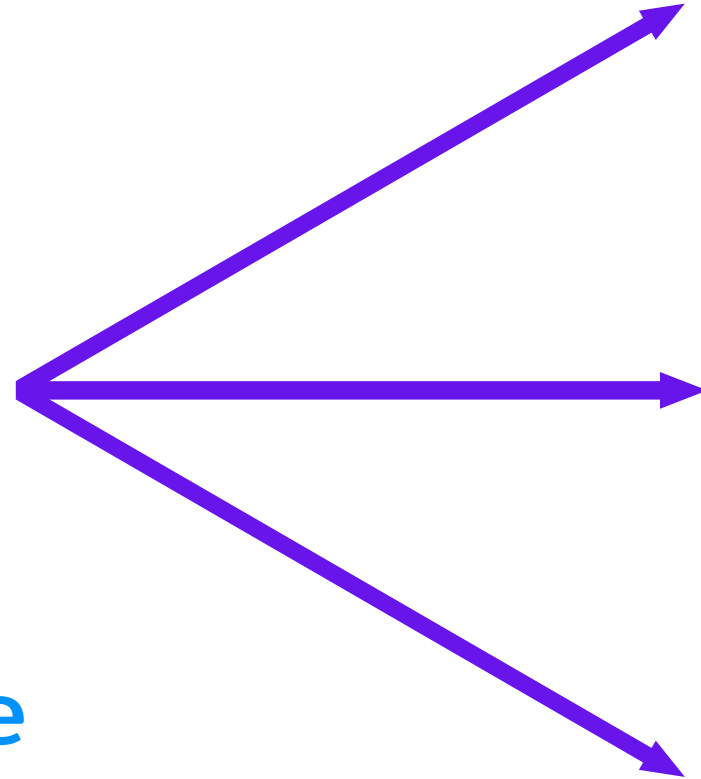


Lipase





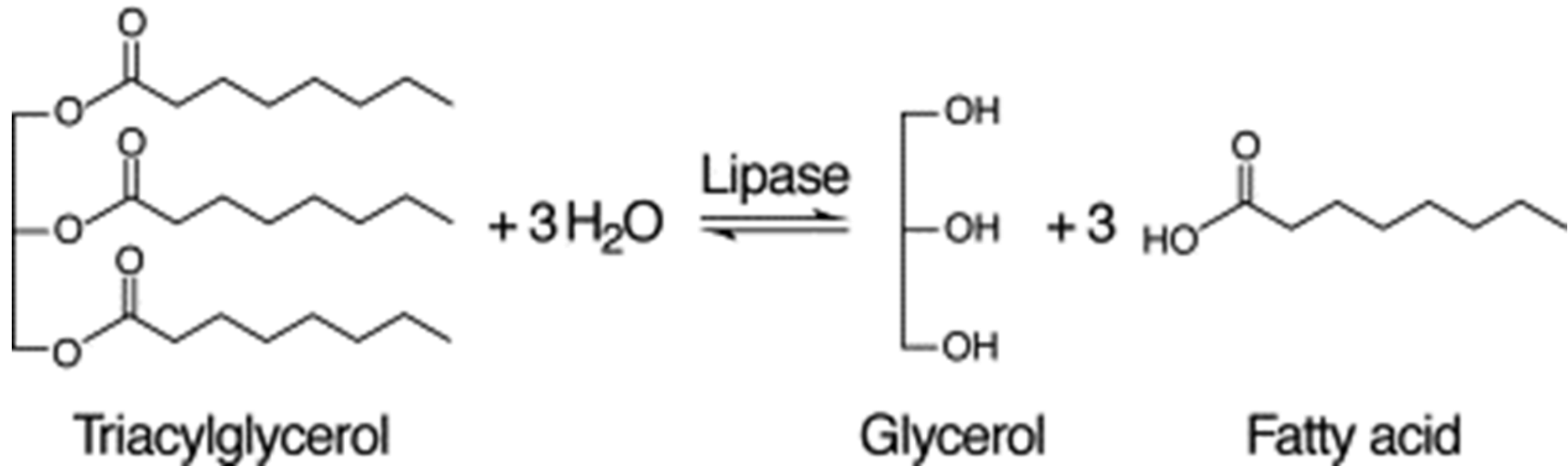
# Application of Lipase



# Introduction

Enzyme from plants

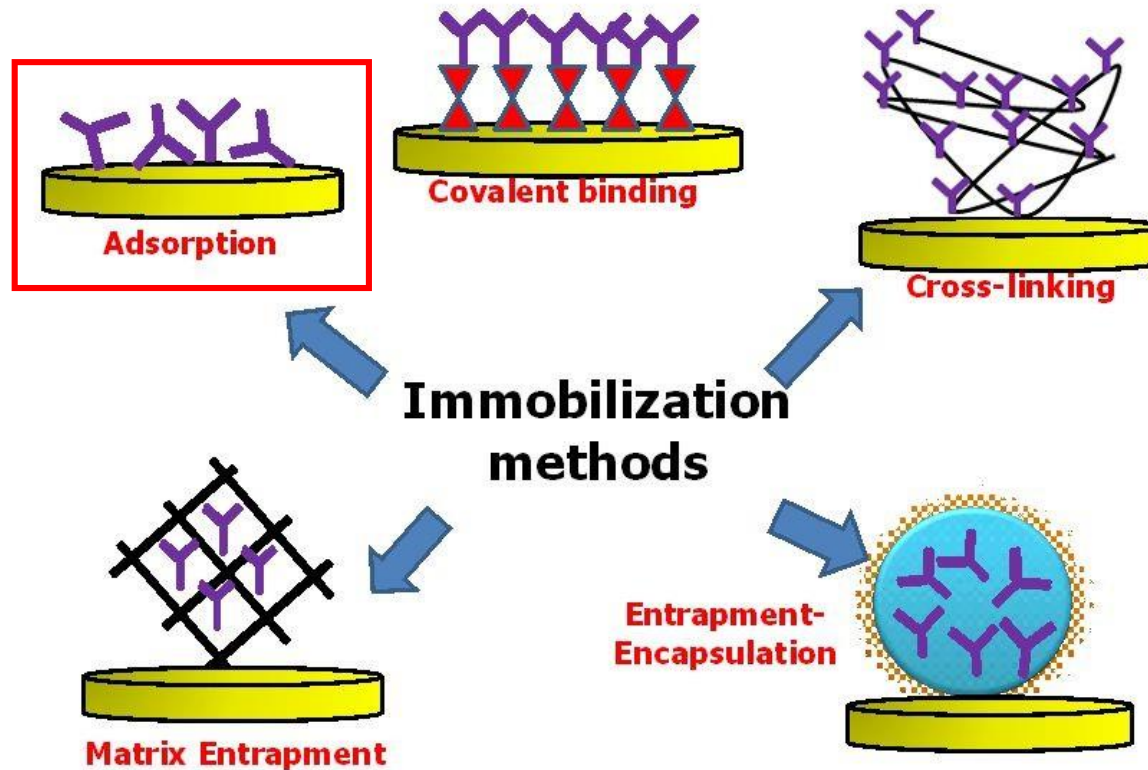
-> Low production cost. We extract from **Pumpkin Seeds**



# Introduction

## Immobilizing enzyme

-> Is to bind enzymes into insoluble solid



> OUR PROJECT <

focuses on

*Chitosan-immobilized*

*lipase!*

## Problem

Which pH and temperature do crude extracted lipase and immobilized lipase have highest activity?

## Hypothesis

The immobilized enzyme is less effective, but it can withstand more severe conditions Ex. pH, Temperature

# Objectives

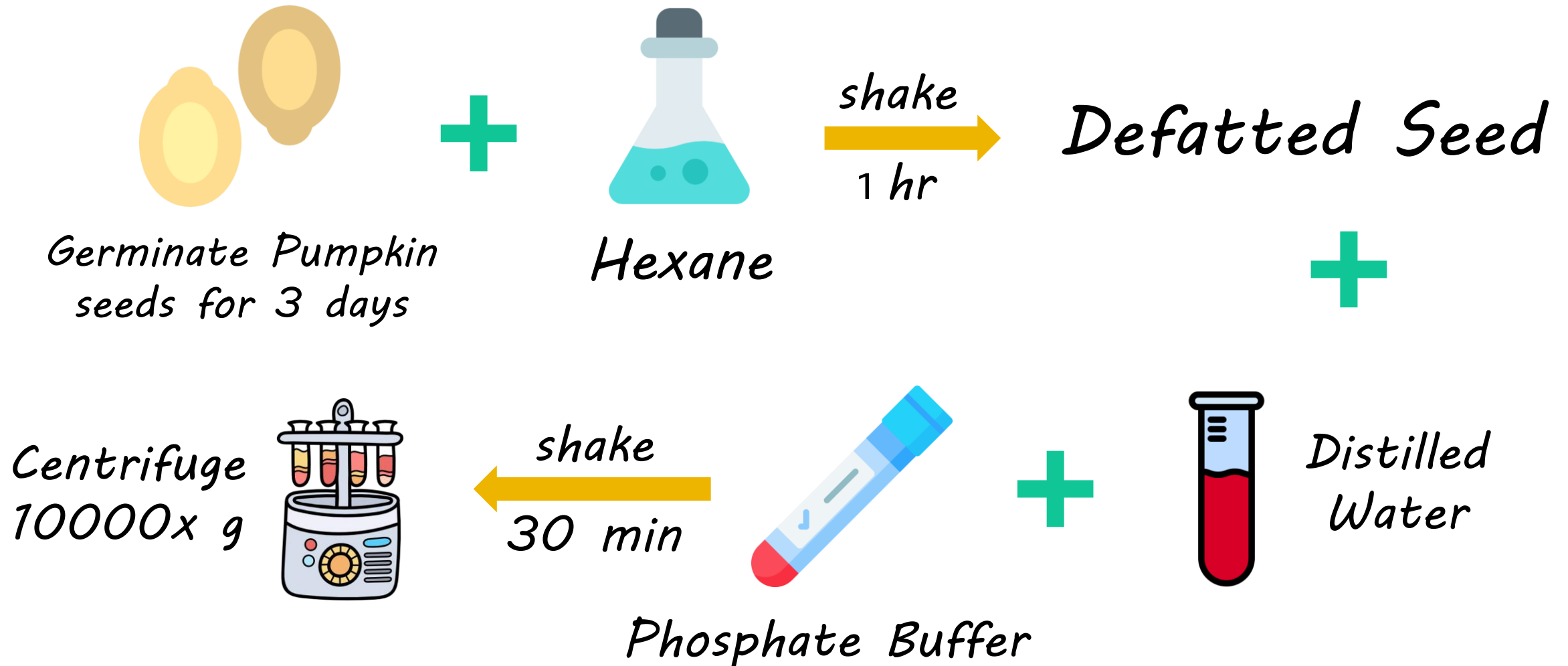
- To extract lipase from germinated pumpkin seeds
- To study lipase activity at various pH and temperatures
- To study properties of lipase and immobilized lipase

## Project scope

- We extract the enzyme from germinated pumpkin seeds
- We use physical adsorption method on Chitosan

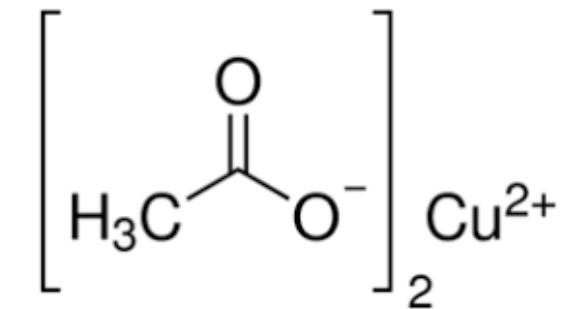


# Part 1: Lipase Extraction

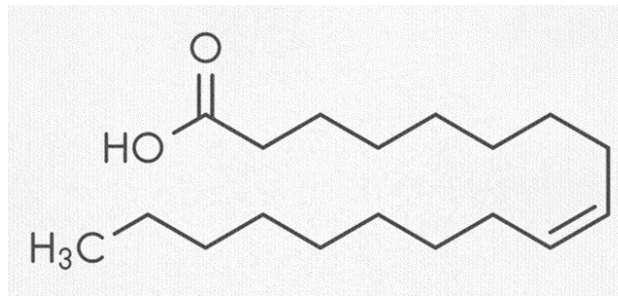


# Part 2 : Lipase Activity Assay

## Copper Acetate Test



*Copper(II) Acetate*

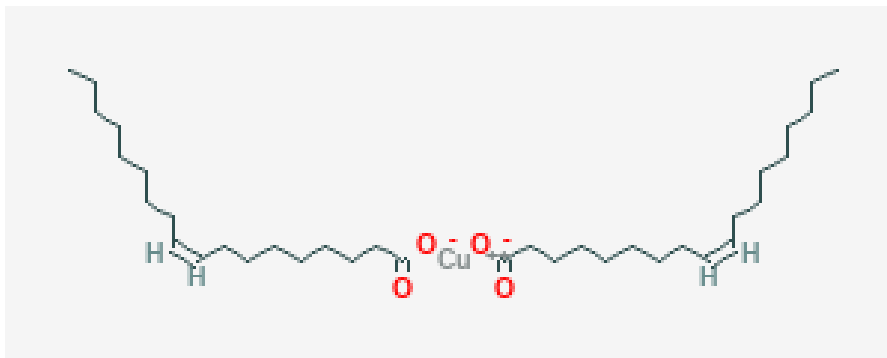


*Oleic Acid*



*Copper(II)  
Oleate  
in Hexane*

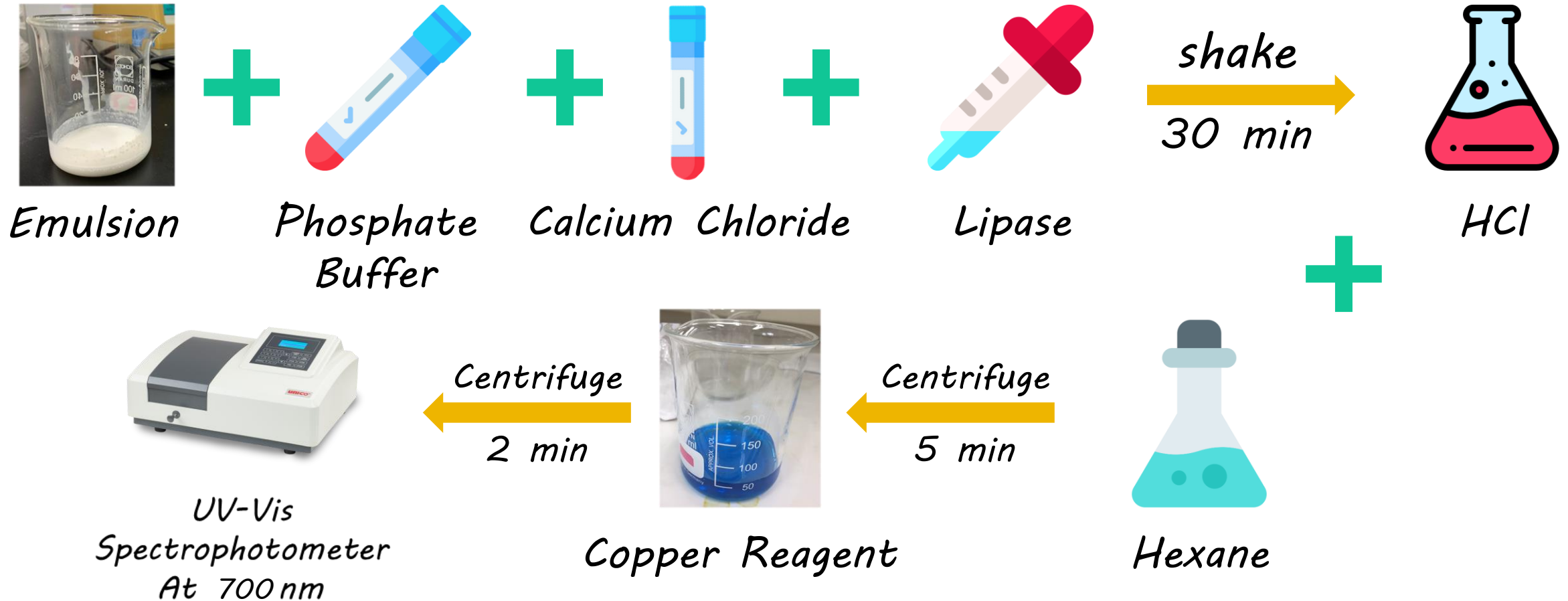
*Copper(II)  
Acetate  
in Water*



*Copper(II) Oleate  
Stable & Clear*

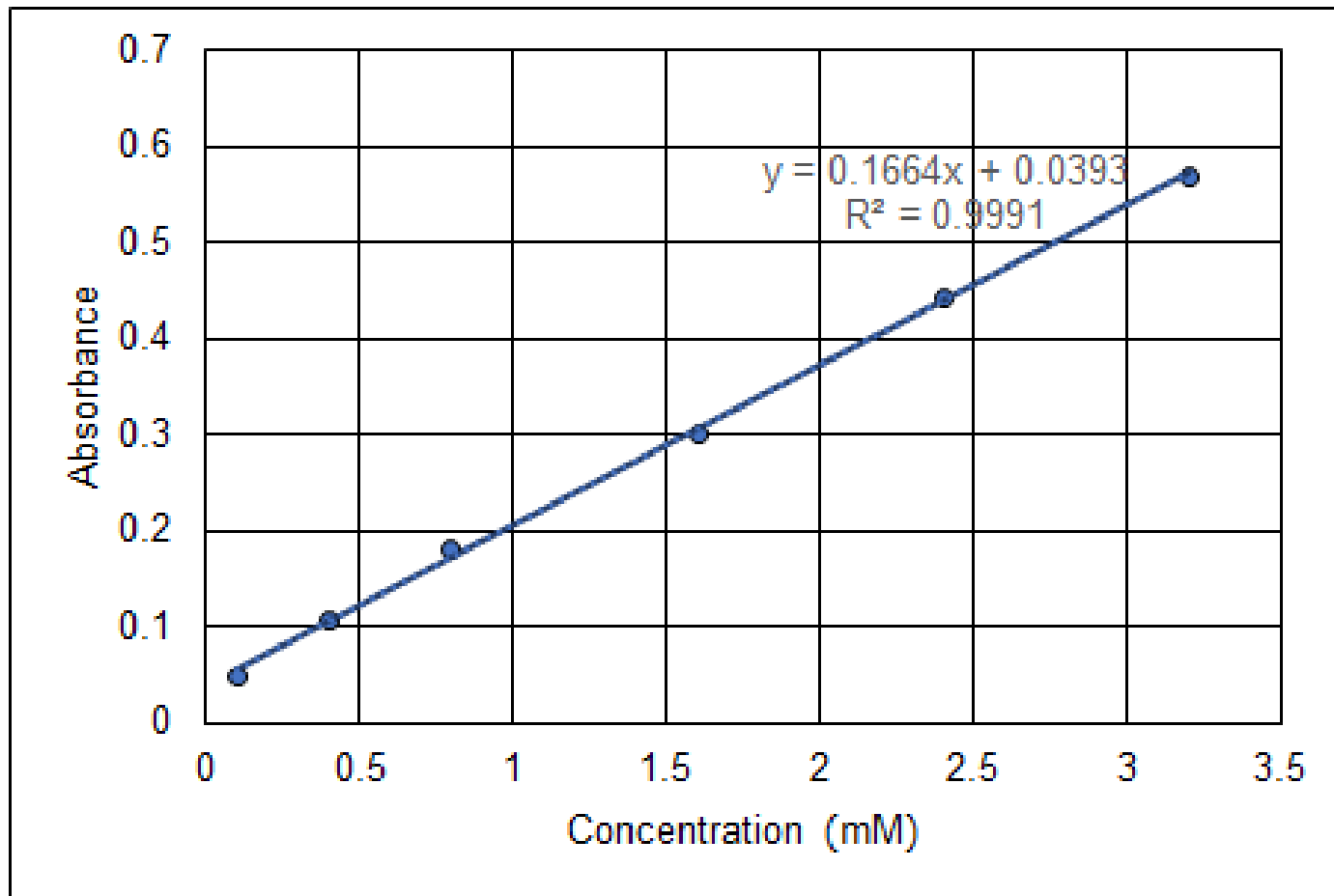
# Part 2 : Lipase Activity Assay

## Activity Assay Process



# Part 2 : Lipase Activity Assay

## Standard Graph of Oleic acid



# Part 3 : Immobilization on Chitosan



*Lipase*

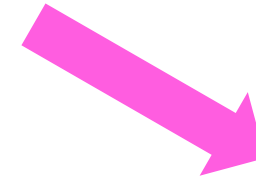


*Chitosan*



*Phosphate  
Buffer*

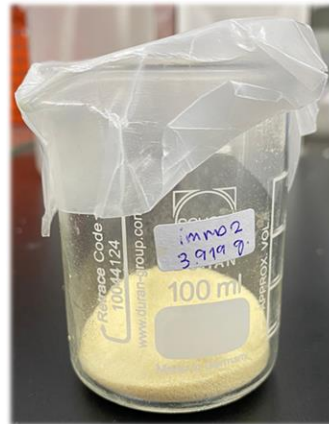
*Stir at 350rpm  
for 7-8 hr*



*Filter and Rinse with Water*



*Dry at 50°C for 12 hr*





# Part 4 : Protein Assay

## Lowry Protein Assay

*Step 1: Protein +  $\text{Cu}^{2+}$   $\rightarrow$   $\text{Cu}^{2+}$ -peptide bond complex (purple)*

*Step 2: Folin reagent +  $\text{Cu}^{2+}$ -peptide bond complex  $\rightarrow$  Reduced folin reagent (blue)*

# Part 4 : Protein Assay

## Lowry Protein Assay

Step 1: Protein +  $\text{Cu}^{2+}$   $\rightarrow$   $\text{Cu}^{2+}$ -peptide bond complex (purple)

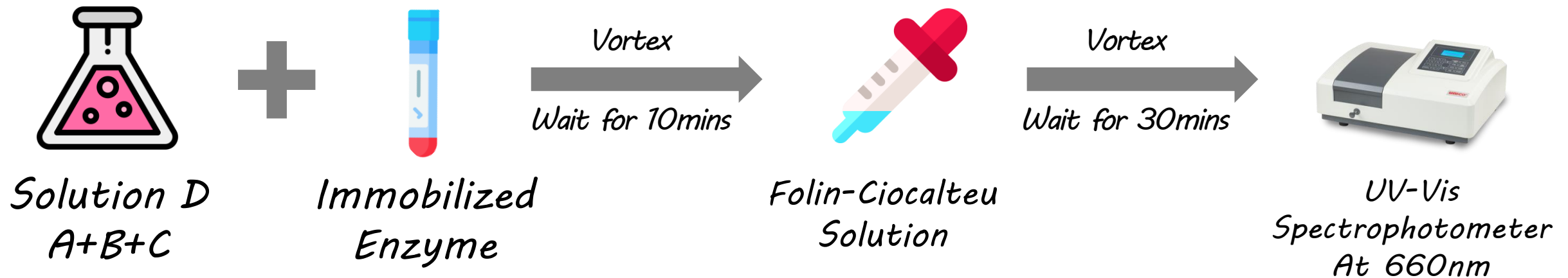
Step 2: Folin reagent +  $\text{Cu}^{2+}$ -peptide bond complex  $\rightarrow$  Reduced folin reagent (blue)

Solution  
Color

*More sensitive than biuret = More accuracy*

# Part 4 : Protein Assay

## Protein Assay Method



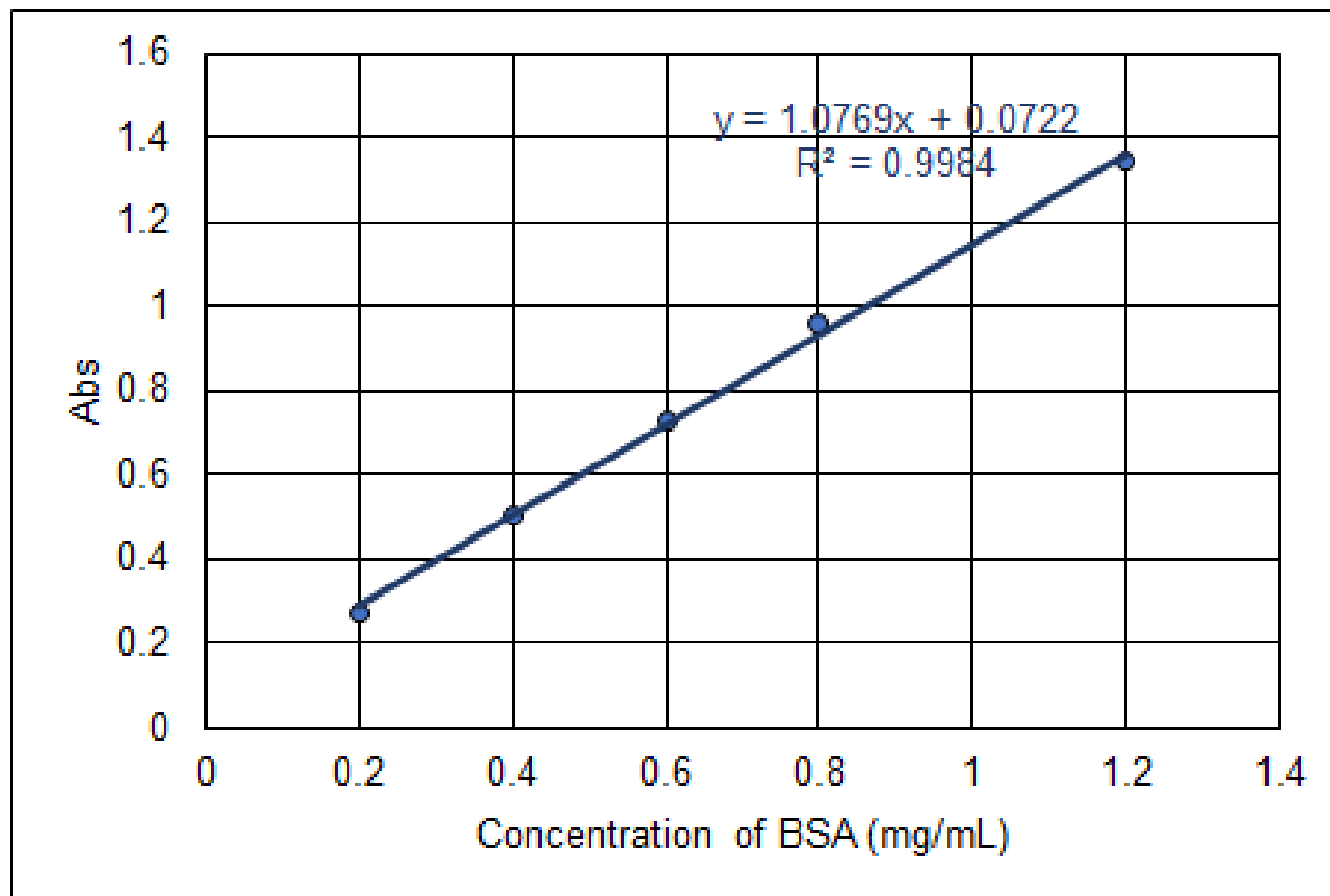
A : Copper(II) Sulfate 0.1 M

B : Sodium Tartrate 0.1 M

C : Sodium Hydroxide 0.1 M + Sodium Carbonate 0.2 M

# Part 4 : Protein Assay

## Standard Graph of BSA



# Data Analysis

## Unit of Lipase

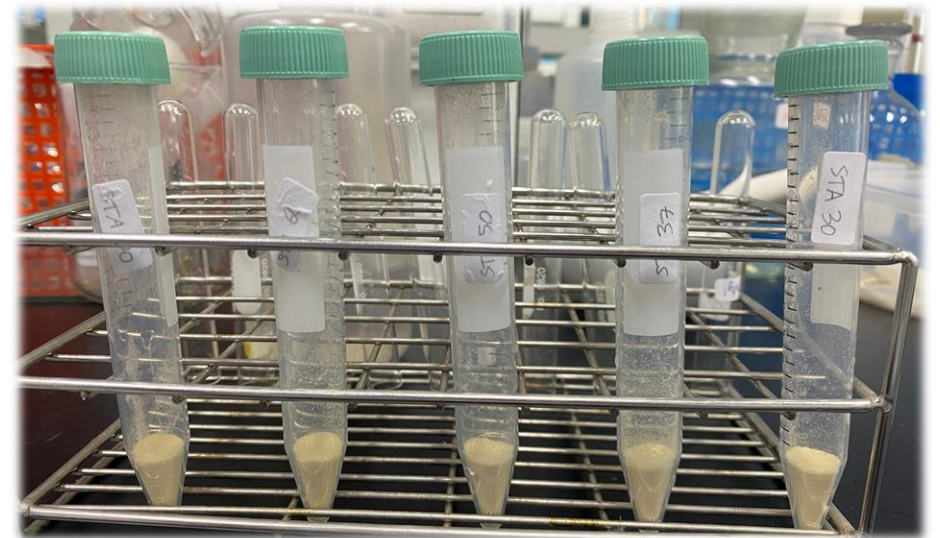
1 unit = quantity of enzyme that liberates the equivalent of 1 $\mu$ mol of free fatty acid per minute

$$\text{Unit} = \frac{C \times V}{\text{min} \times 1000}$$

C = Concentration of Oleic acid from the incubation ( $\mu$ M)

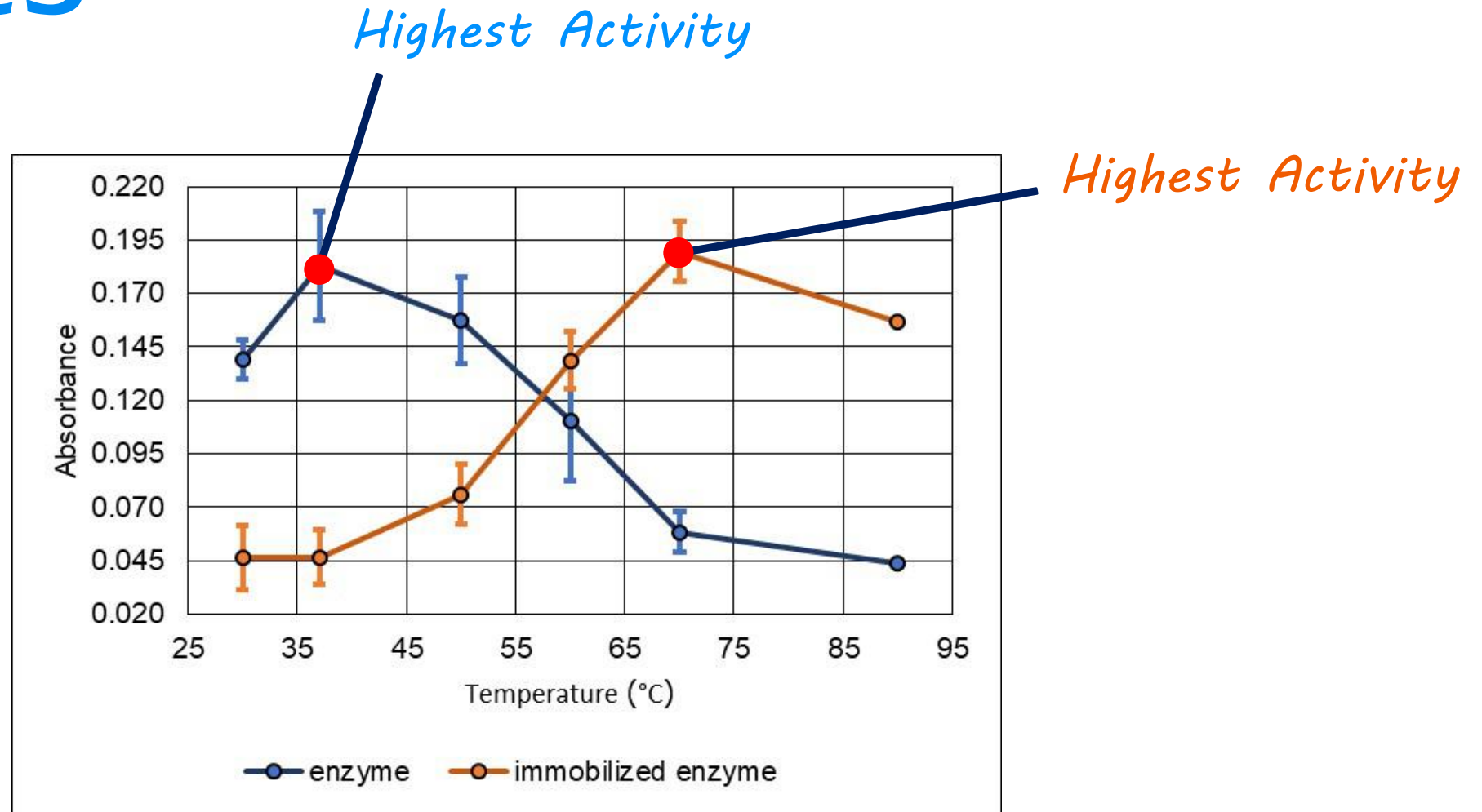
V = Volume of substance used during incubation (mL)

min = Incubation time



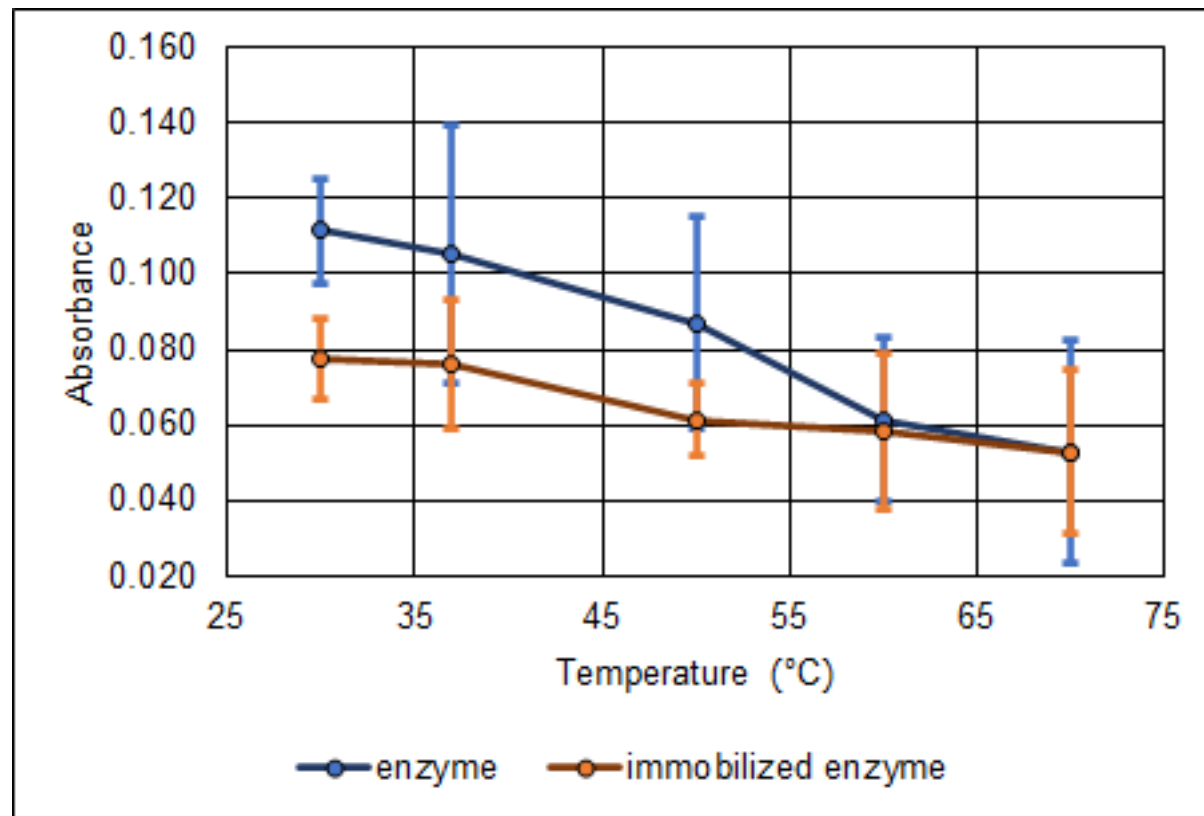


# Results



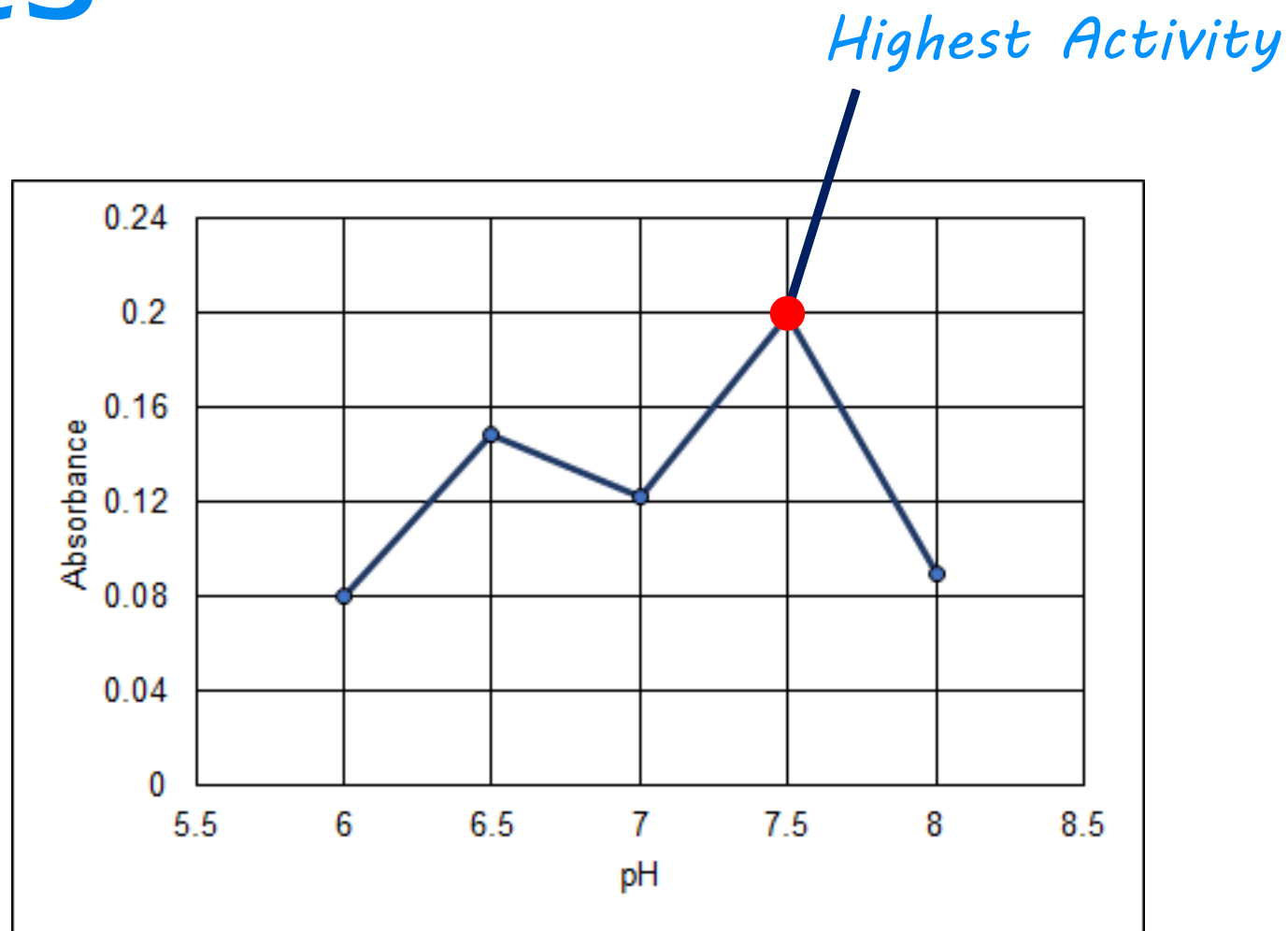
*Absorbance at different incubating temperature*

# Results



*Absorbance after putting the enzyme in different water temperature for 1hr*

# Results



*Absorbance in different pH of Phosphate Buffer during incubation*

# Conclusion

- Crude extracted lipase from germinated pumpkin seeds have 0.559 Unit of lipase per gram of peeled pumpkin seeds
- Crude extracted enzyme has an optimal pH, temperature at 7.5, 37 °C respectively and immobilized enzyme has an optimum temperature at 70 °C
- Chitosan can bind 22.4274 % of protein and has high activity

# References

Barros, M.; Fleuri, L. F.; Macedo, G. A. Seed Lipases: Sources, Applications, and Properties - A Review. Brazilian Journal of Chemical Engineering. 2010, 27(1), 15-29.

Balan, A; Ibrahim, D.; Rahim, R. A.; Rashid, F. A. A. Purification and Characterization of a Thermostable Lipase From *Geobacillus Thermodenitrificans* IBRL-nra. Enzyme Research. 2012, 1-7.

Eze, S. O. O.; Chilaka, F. C. Lipolytic Activities in Some Species of Germinating Cucurbitaceae: *Cucumeropsis manii* Naud, *Colocynthis vulgaris* L. and *Cucurbita moschata* Schrad. World Applied Sciences Journal. 2010, 10(10), 1225-123.

Avramiuc, M. Comparative Study on the Lipase Activity from Plant Sources, Under Various Conditions of pH, Temperature, and Substrate. Journal of Faculty of Food Engineering. 2016, 1, 21-28.

Eze, S. O. O.; Eczema, B. O. Purification and Characterization of Lipase (EC-3.1.1.3) from the Seeds of *Cucumeropsis mannii* (White Melon). Thai Journal of Agricultural Science. 2012, 45(2), 115-120.



*Thank You*